

# **Product Data Sheet**

# Lysosome Deep Red Probes

Catalog #	Source	e Reactivity	Applications
CRG1109			IF
Description		Deep Red Fluorescen	t acidotropic probes for labeling and tracking acidic organelles
		in live cells	
Specificity		Weakly basic amines	selectively accumulate in cellular compartments with low
		internal pH and can b	e used to investigate the biosynthesis and pathogenesis of
		lysosomes. The Lysos	ome Deep Red Probes are fluorescent acidotropic probes for
		labeling and tracking	acidic organelles in live cells. These probes have several
		important features, ir	ncluding high selectivity for acidic organelles and effective
		labeling of live cells a	t nanomolar concentrations.
Form		Liquid	
Application		The Lysosome Deep F	Red Probes, which consist of a fluorophore linked to a weak
		base that is only part	ially protonated at neutral pH, are freely permeant to cell
		membranes and typic	cally concentrate in spherical organelles. Their mechanism of
		retention has not bee	en firmly established but is likely to involve protonation and
		retention in the mem	branes of the organelles, although staining is generally not
		reversed by subseque	ent treatment of the cells with weakly basic cell-permeant
		compounds. Note that	at in Lysosome Deep Red Probes stained cells, the lysosomal
		fluorescence may cor	nstitute only a small portion of total cellular fluorescence,
		making it difficult to o	quantitate the number of lysosomes by flow cytometry or
		fluorometry.	
<b>Directions for</b>	Use	1. Prepare the workir	ng solution
		Add Lysosome Deep I	Red Probes to the cell culture medium at the ratio of 1:10,000 -
		1:20,000. The final co	ncentration is 50-100 nM. The working solution can be
		warmed at 37°C befo	re use.
		2. Label the lysosome	25
		For adherent cells, di	scard the cell medium and wash with 1 x PBS. Adding the
Directions for	Use	membranes and typic retention has not been retention in the mem- reversed by subseque compounds. Note that fluorescence may cor- making it difficult to of fluorometry. 1. Prepare the workin Add Lysosome Deep H 1:20,000. The final co- warmed at 37°C befor 2. Label the lysosome For adherent cells, different	cally concentrate in spherical organelles. Their mechanism of en firmly established but is likely to involve protonation and branes of the organelles, although staining is generally not ent treatment of the cells with weakly basic cell-permeant at in Lysosome Deep Red Probes stained cells, the lysosomal institute only a small portion of total cellular fluorescence, quantitate the number of lysosomes by flow cytometry or ang solution Red Probes to the cell culture medium at the ratio of 1:10,000 - oncentration is 50-100 nM. The working solution can be re use.

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working solution, and incubate the cells at 37°C for 30 mins to 2 hours. Discard the working solution, wash with 1 x PBS for three times, and then photographed under a fluorescence microscope.

For suspension cells, centrifuge to obtain a cell pellet and aspirate the supernatant. Resuspend the cells gently in probe-containing medium, and incubate the cells at 37°C for 30 mins to 2 hours. Centrifuge to obtain a cell pellet and aspirate the supernatant, wash with 1 x PBS for three times, and then photographed under a fluorescence microscope.

#### Notice

1. If the staining effect is not good, the concentration of the probe in the working fluid can increase the concentration of the working solution , or can extend the reaction time.

2. In order to reduce the background, pleasee use lower concentration probes.

3. Take photos quickly, because the dye is easy to quench.

**Storage/Stability** Store at -20 °C in the dark for 6 months.

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