

Histamine Dehydrogenase Microplate Assay Kit User Manual

Catalog # CAK1307

(Version 1.1A)

Detection and Quantification of Histamine Dehydrogenase (HDH)
Activity in Tissue extracts, Cell lysate, Cell culture media and Other biological fluids Samples.

For research use only. Not for diagnostic or therapeutic procedures.



I. INTRODUCTION	2
II. KIT COMPONENTS	3
III. MATERIALS REQUIRED BUT NOT PROVIDED	3
IV. SAMPLE PREPARATION	4
V. ASSAY PROCEDURE	5
VI. CALCULATION	6
VII. TYPICAL DATA	7
VIII. TECHNICAL SUPPORT	7
IX NOTES	7



I. INTRODUCTION

Histamine dehydrogenase is a homodimeric enzyme and catalyzes oxidative deamination of histamine. Histamine dehydrogenase can be isolated from cultures of Nocardiodes simplex grown on histamine as the sole nitrogen source. The enzyme is a homodimer of a ~76kDa subunit responsible for catalyzing the oxidative deamination of histamine to give imidazole acetaldehyde (Scheme 1), where the enzyme exhibits remarkable selectivity for histamine, thus showing great potential for use in biosensors. HADH was originally classified as a quinone-containing amine dehydrogenase. Later studies revealed that HADH is a homologue of TMADH, and dimethylamine dehydrogenase (DMADH) from M. methylotrophus shares 40% sequence identity and 56% similarity with both proteins.

Histamine Dehydrogenase Microplate Assay Kit is a sensitive assay for determining histamine dehydrogenase activity in various samples. Histamine is hydrolyzed by histamine dehydrogenase. The intensity of product color, measured at 450 nm is directly proportional to histamine dehydrogenase activity in the sample.



II. KIT COMPONENTS

Component	Volume	Storage
96-Well Microplate	1 plate	
Assay Buffer	30 ml x 4	4 °C
Reaction Buffer	15 ml x 1	4 °C
Substrate	Powder x 1	4 °C
Dye Reagent A	Powder x 1	4 °C
Dye Reagent B	1 ml x 1	4 °C
Standard	Powder x 1	4 °C
Positive Control	Powder x 1	-20 °C
Technical Manual	1 Manual	

Note:

Dye Reagent A: add 1 ml distilled water to dissolve before use, mix. Store at -20°C for a month.

Substrate: add 1 ml Reaction Buffer to dissolve before use, mix. Store at -20°C for a month.

Standard: add 1 ml distilled water to dissolve, mix; the concentration will be 1 mmol/L. Store at -20°C for a month.

Positive Control: add 0.5 ml Assay Buffer to dissolve before use; store at -80 °C for a month after reconstitution.



III. MATERIALS REQUIRED BUT NOT PROVIDED

- 1. Microplate reader to read absorbance at 450 nm
- 2. Distilled water
- 3. Pipettor, multi-channel pipettor
- 4. Pipette tips
- 5. Mortar
- 6. Centrifuge
- 7. Timer

IV. SAMPLE PREPARATION

1. For cell and bacteria samples

Collect cell or bacteria into centrifuge tube, discard the supernatant after centrifugation, add 1 ml Assay buffer for 5×10^6 cell or bacteria, sonicate (with power 20%, sonicate 3s, interval 10s, repeat 30 times); centrifuged at 8000g 4 °C for 10 minutes, take the supernatant into a new centrifuge tube and keep it on ice for detection.

2. For tissue samples

Weigh out 0.1 g tissue, homogenize with 1 ml Assay buffer on ice, centrifuged at 8000g 4 °C for 10 minutes, take the supernatant into a new centrifuge tube and keep it on ice for detection.

3. For liquid samples

Liquid samples can be used directly.



V. ASSAY PROCEDURE

Warm all regents to room temperature before use.

Add following reagents into the microplate:

Reagent	Sample	Control	Standard	Blank	Positive
					Control
Substrate	170 μΙ	170 μΙ	170 μΙ	170 μΙ	170 μΙ
Sample	10 μΙ				
Assay Buffer		10 μΙ			
Positive Control					10 μΙ
Standard			10 μΙ		
Distilled water				10 μΙ	
Dye Reagent A	10 μΙ	10 μΙ	10 μΙ	10 μΙ	10 μΙ
Dye Reagent B	10 μΙ	10 μΙ	10 μΙ	10 μΙ	10 μΙ

Mix, incubate at room temperature for 5 minutes, measured at 450 nm and record the absorbance.

Note:

- 1) Perform 2-fold serial dilutions of the top standards to make the standard curve.
- 2) The concentrations can vary over a wide range depending on the different samples. For unknown samples, we recommend doing a pilot experiment & testing several doses to ensure the readings are within the standard curve range.
- 3) Reagents must be added step by step, can not be mixed and added together.



VI. CALCULATION

Unit Definition: One unit of HDH activity is defined as the amount of enzyme which reduces 1 μ mol of H⁺ per min at 37°C.

1. According to the volume of sample

$$\begin{split} \text{HDH (}\mu\text{mol/ml)} &= \left(C_{\text{Standard}} \times V_{\text{Standard}} \right) \times \left(\text{OD}_{\text{Sample}} - \text{OD}_{\text{Control}} \right) / \left(\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}} \right) / \\ & V_{\text{Sample}} / T \\ &= 0.2 \times \left(\text{OD}_{\text{Sample}} - \text{OD}_{\text{Control}} \right) / \left(\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}} \right) \end{split}$$

2. According to the weight of sample

$$\begin{split} \text{HDH (}\mu\text{mol/g}) &= \left(\text{C}_{\text{Standard}} \times \text{V}_{\text{Standard}}\right) \times \left(\text{OD}_{\text{Sample}} - \text{OD}_{\text{Control}}\right) / \left(\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}}\right) / \left(\text{W} \times \text{V}_{\text{Sample}} / \text{V}_{\text{Assay}}\right) / T \\ &= 0.2 \times \left(\text{OD}_{\text{Sample}} - \text{OD}_{\text{Control}}\right) / \left(\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}}\right) / \text{W} \end{split}$$

C_{Protein}: the protein concentration, mg/ml;

 $C_{Standard}$: the standard concentration, 1 mmol/L = 1 μ mol/ml;

W: the weight of sample, g;

V_{Sample}: the volume of sample, 0.01 ml;

V_{Standard}: the volume of standard, 0.01 ml;

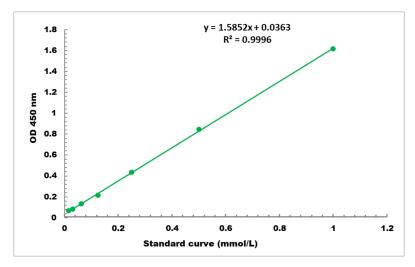
V_{Assay}: the volume of Assay buffer, 1 ml;

T: the reaction time, 5 minutes.

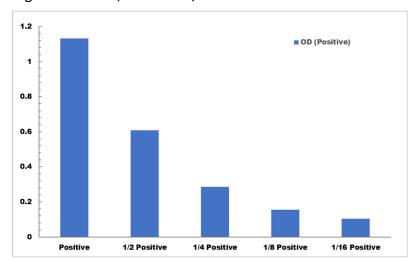


VII. TYPICAL DATA

The standard curve is for demonstration only. A standard curve must be run with each assay.



Detection Range: 0.01 mmol/L - 1 mmol/L



Positive Control reaction in 96-well plate assay with decreasing the concentration

VIII. TECHNICAL SUPPORT

For troubleshooting, information or assistance, please go online to www.cohesionbio.com or contact us at techsupport@cohesionbio.com

IX. NOTES