



# **3-Phosphoglycerate Kinase Microplate Assay Kit User Manual**

**Catalog # CAK1287**

(Version 1.1A)

Detection and Quantification of 3-Phosphoglycerate Kinase (PGK)  
Activity in Serum, Plasma, Tissue extracts, Cell lysate, Cell culture  
media, Other biological fluids Samples.

**For research use only. Not for diagnostic or therapeutic procedures.**

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## I. INTRODUCTION

3-Phosphoglycerate kinase (EC 2.7.2.3) (PGK) is an enzyme that catalyzes the reversible transfer of a phosphate group from 1,3-bisphosphoglycerate (1,3-BPG) to ADP producing 3-phosphoglycerate (3-PG) and ATP. Like all kinases it is a transferase. PGK is a major enzyme used in glycolysis, in the first ATP-generating step of the glycolytic pathway. In gluconeogenesis, the reaction catalyzed by PGK proceeds in the opposite direction, generating ADP and 1,3-BPG.

3-Phosphoglycerate Kinase Microplate Assay Kit provides a simple and sensitive method for monitoring 3-Phosphoglycerate Kinase activity in various samples. In this assay, 3-Phosphoglycerate Kinase catalyzes conversion of 3-phosphoglycerate to 1,3-diphosphoglycerate and an intermediate, which reacts with a developer to form a colored product that absorbs maximally at 492 nm.

## II. KIT COMPONENTS

Component	Volume	Storage
96-Well Microplate	1 plate	
Assay Buffer	30 ml x 4	4 °C
Reaction Buffer	10 ml x 1	4 °C
Substrate	Powder x 1	-20 °C
Enzyme	Powder x 1	-20 °C
Dye Reagent A	Powder x 1	4 °C
Dye Reagent B	1 ml x 1	4 °C
Standard	Powder x 1	-20 °C
Positive Control	100 µl x 1	4 °C
Plate Adhesive Strips	3 Strips	
Technical Manual	1 Manual	

### Note:

**Substrate:** add 1 ml Reaction Buffer to dissolve before use, store at 4 °C for 1-2 weeks after reconstitution.

**Enzyme:** add 1 ml Reaction Buffer to dissolve before use, store at -80 °C for 1 month after reconstitution.

**Dye Reagent A:** add 9 ml distilled water to dissolve before use, mix, store at 4°C for 1 month after reconstitution.

**Standard:** add 1 ml distilled water to dissolve before use; then add 0.15 ml into 0.85 ml distilled water, the concentration will be 300 µmol/L, store at 4°C for 1-2 weeks after reconstitution.

### **III. MATERIALS REQUIRED BUT NOT PROVIDED**

1. Microplate reader to read absorbance at 492 nm
2. Distilled water
3. Pipettor, multi-channel pipettor
4. Pipette tips
5. Mortar
6. Centrifuge
7. Timer

### **IV. SAMPLE PREPARATION**

1. For cell and bacteria samples

Collect cell or bacteria into centrifuge tube, discard the supernatant after centrifugation, add 1 ml Assay Buffer for  $5 \times 10^6$  cell or bacteria, sonicate (with power 20%, sonicate 3s, interval 10s, repeat 30 times); centrifuged at 8000g 4 °C for 10 minutes, take the supernatant into a new centrifuge tube and keep it on ice for detection.

2. For tissue samples

Weigh out 0.1 g tissue, homogenize with 1 ml Assay Buffer on ice, centrifuged at 8000g 4 °C for 10 minutes, take the supernatant into a new centrifuge tube and keep it on ice for detection.

3. For serum or plasma samples

Detect directly.

## V. ASSAY PROCEDURE

Warm the all reagents to room temperature before use.

Add following reagents into the microplate:

Reagent	Sample	Control	Standard	Blank	Positive Control
Reaction Buffer	70 µl	70 µl	--	--	70 µl
Coenzyme	10 µl	10 µl	--	--	10 µl
Substrate	10 µl	10 µl	--	--	10 µl
Sample	10 µl	--	--	--	--
Positive Control	--	--	--	--	10 µl
Standard	--	--	100 µl	--	--
Distilled water	--	10 µl	--	100 µl	--
Dye Reagent A	90 µl	90 µl	90 µl	90 µl	90 µl
Dye Reagent B	10 µl	10 µl	10 µl	10 µl	10 µl
Mix, cover the plate adhesive strip, put the plate into the convection oven, incubate at 37 °C for 5 minutes, measured at 492 nm and record the absorbance.					

### Note:

- 1) Perform 2-fold serial dilutions of the top standards to make the standard curve.
- 2) For unknown samples, we recommend doing a pilot experiment & testing several doses to ensure the readings are within the standard curve range. If the enzyme activity is lower, please add more sample into the reaction system; or increase the reaction time; if the enzyme activity is higher, please dilute the sample, or decrease the reaction time.
- 3) Reagents must be added step by step, can not be mixed and added together.

## VI. CALCULATION

**Unit Definition:** one unit of PGK activity is defined as the enzyme oxidize 1  $\mu\text{mol}$  NADH per minute.

### 1. According to the protein concentration of sample

$$\begin{aligned}\text{PGK (U/mg)} &= (C_{\text{Standard}} \times V_{\text{Standard}}) \times (\text{OD}_{\text{Control}} - \text{OD}_{\text{Sample}}) / (\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}}) / (V_{\text{Sample}} \\ &\quad \times C_{\text{Protein}}) / T \\ &= 0.6 \times (\text{OD}_{\text{Control}} - \text{OD}_{\text{Sample}}) / (\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}}) / C_{\text{Protein}}\end{aligned}$$

### 2. According to the weight of sample

$$\begin{aligned}\text{PGK (U/g)} &= (C_{\text{Standard}} \times V_{\text{Standard}}) \times (\text{OD}_{\text{Control}} - \text{OD}_{\text{Sample}}) / (\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}}) / (V_{\text{Sample}} \times \\ &\quad W / V_{\text{Assay}}) / T \\ &= 0.6 \times (\text{OD}_{\text{Control}} - \text{OD}_{\text{Sample}}) / (\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}}) / W\end{aligned}$$

### 3. According to the quantity of cells or bacteria

$$\begin{aligned}\text{PGK (U/10}^4\text{)} &= (C_{\text{Standard}} \times V_{\text{Standard}}) \times (\text{OD}_{\text{Control}} - \text{OD}_{\text{Sample}}) / (\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}}) (V_{\text{Sample}} \\ &\quad \times N / V_{\text{Assay}}) / T \\ &= 0.6 \times (\text{OD}_{\text{Control}} - \text{OD}_{\text{Sample}}) / (\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}}) / N\end{aligned}$$

### 4. According to the volume of sample

$$\begin{aligned}\text{PGK (U/ml)} &= (C_{\text{Standard}} \times V_{\text{Standard}}) \times (\text{OD}_{\text{Control}} - \text{OD}_{\text{Sample}}) / (\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}}) / V_{\text{Sample}} / \\ &\quad T \\ &= 0.6 \times (\text{OD}_{\text{Control}} - \text{OD}_{\text{Sample}}) / (\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}})\end{aligned}$$

$C_{\text{Standard}}$ : the standard concentration,  $600 \mu\text{mol/L} = 0.6 \mu\text{mol/ml}$ ;

$V_{\text{Standard}}$ : the volume of standard,  $100 \mu\text{l} = 0.1 \text{ ml}$ ;

$C_{\text{Protein}}$ : the protein concentration,  $\text{mg/ml}$ ;

$W$ : the weight of sample,  $\text{g}$ ;

$N$ : the quantity of cell or bacteria,  $N \times 10^4$ ;

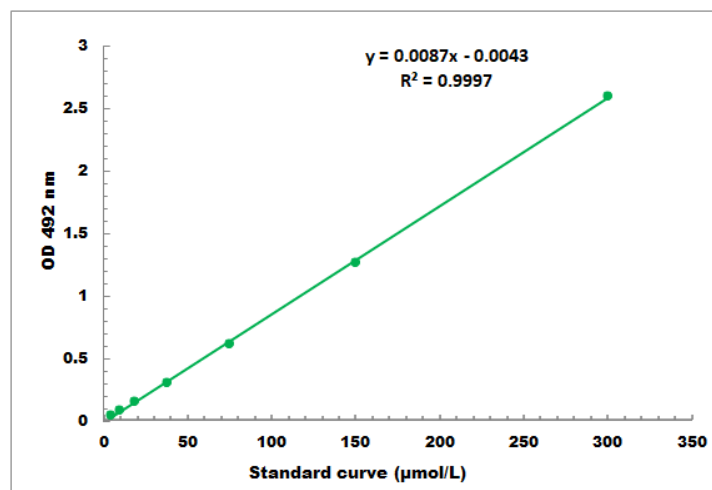
$V_{\text{Sample}}$ : the volume of sample,  $0.01 \text{ ml}$ ;

$V_{\text{Assay}}$ : the volume of Assay buffer,  $1 \text{ ml}$ ;

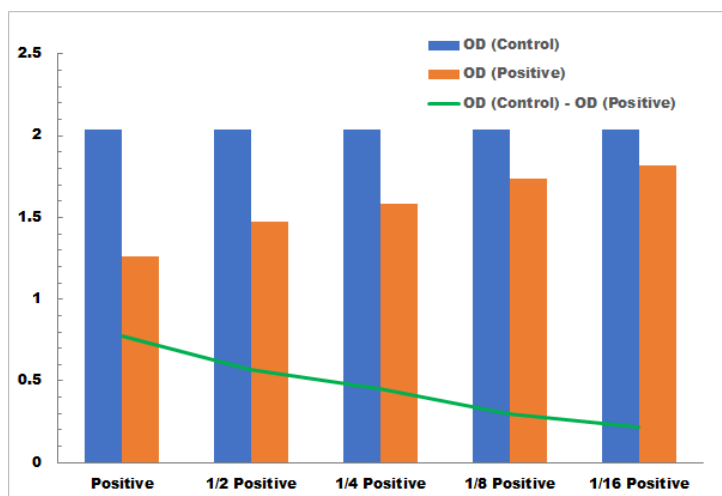
$T$ : the reaction time,  $5 \text{ minutes}$ .

## VII. TYPICAL DATA

The standard curve is for demonstration only. A standard curve must be run with each assay.



Detection Range: 3 μmol/L - 300 μmol/L



Positive Control reaction in 96-well plate assay with decreasing the concentration

## VIII. TECHNICAL SUPPORT

For troubleshooting, information or assistance, please go online to [www.cohesionbio.com](http://www.cohesionbio.com) or contact us at [techsupport@cohesionbio.com](mailto:techsupport@cohesionbio.com)

## IX. NOTES