

Urea Colorimetric Microplate Assay Kit User Manual

Catalog # CAK1160

(Version 1.5D)

Detection and Quantification of UreaContentin Urine, Serum, Tissue extracts, Cell lysate, Cell culture, Other biological fluids mediaSamples.

For research use only. Not for diagnostic or therapeutic procedures.



| I. INTRODUCTION | 2 |
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| II. KIT COMPONENTS | 3 |
| III. MATERIALS REQUIRED BUT NOT PROVIDED | 3 |
| IV. SAMPLE PREPARATION | 4 |
| V. ASSAY PROCEDURE | 5 |
| VI. CALCULATION | 6 |
| VII. TYPICAL DATA | 7 |
| VIII. TECHNICAL SUPPORT | 7 |
| IX. NOTES | 7 |



I. INTRODUCTION

Urea is primarily produced in the liver and secreted by the kidneys. Urea is the major end product of protein catabolism in animals. It is the primary vehicle for removal of toxic ammonia from the body. Urea determination is very useful for the medical clinician to assess kidney function of patients. In general, increased urea levels are associated with nephritis, renal ischemia, urinary tract obstruction, and certain extrarenal diseases, e.g., congestive heart failure, liver diseases and diabetes.

Decreased levels indicate acute hepatic insufficiency or may result from overvigorous parenteral fluid therapy.

UreaColorimetric Microplate Assay Kit is designed to measure urea directly in biological samples without any pretreatment. The intensity of the color, measured at 620nm, is directly proportional to the urea concentration in the sample.



II.KIT COMPONENTS

| Component | Volume | Storage |
|------------------------|------------|---------|
| 96-Well Microplate | 1 plate | |
| Enzyme | Powder x 1 | -20 °C |
| Reaction Buffer | 10 ml x 1 | 4 °C |
| Dye Reagent I | Powder x 1 | 4 °C |
| Dye Reagent II | Powder x 1 | 4 °C |
| Dye Reagent II Diluent | 3 ml x 1 | 4 °C |
| Standard | Powder x 1 | 4 °C |
| Plate Adhesive Strips | 3 Strips | |
| Technical Manual | 1 Manual | |

Note:

Enzyme: add 1 mlReaction BufferintoEnzymetube to dissolve before use.

Dye Reagent I: add 7 ml distilled water to dissolve before use.

Dye Reagent II: add 3 mlDye Reagent II DiluentintoDye Reagent II,mix before use.

Standard: add 1 ml distilled water to dissolve before use; then add 0.05 ml into 0.95 ml distilled water, mix, the concentration will be 250 mg/L.

III. MATERIALS REQUIRED BUT NOT PROVIDED

- 1. Microplate reader to read absorbance at 620 nm
- 2. Distilled water
- 3. Pipettor, multi-channel pipettor
- 4. Pipette tips
- 5. Mortar
- 6. Centrifuge
- 7. Timer
- 8. Hot air circulation oven



IV. SAMPLE PREPARATION

1. For urine, serum orother biological fluids samples Detect directly.



V. ASSAY PROCEDURE

Add following reagents in the microplate:

| Reagent | Sample | Standard | Blank | | |
|--|--------|----------|-------|--|--|
| Reaction Buffer | 80 μΙ | 80 μΙ | 80 μΙ | | |
| Sample | 10μΙ | | | | |
| Standard | | 10 μΙ | | | |
| Distilled water | | | 10 μΙ | | |
| Enzyme | 10 μΙ | 10 μΙ | 10 μΙ | | |
| Shake and mix, put it into the oven,37°C for 10minutes. | | | | | |
| Dye Reagent I | 70 μΙ | 70 μl | 70 μΙ | | |
| Dye Reagent II | 30 μΙ | 30 μΙ | 30 μΙ | | |
| Shake and mix, put it into the oven,37°C for 30minutes. Then record absorbance | | | | | |
| measured at 620 nm. | | | | | |

Note:

- 1) Perform 2-fold serial dilutions of the top standards to make the standard curve.
- 2) The concentrations can vary over a wide range depending on the different samples. For unknown samples, we recommend doing a pilot experiment & testing several doses to ensure the readings are within the standard curve range.
- 3)Reagents must be added step by step, can not be mixed and added together.



VI. CALCULATION

1. According to the volume of sample

C_{Standard}: the standard concentration, 250 mg/L;

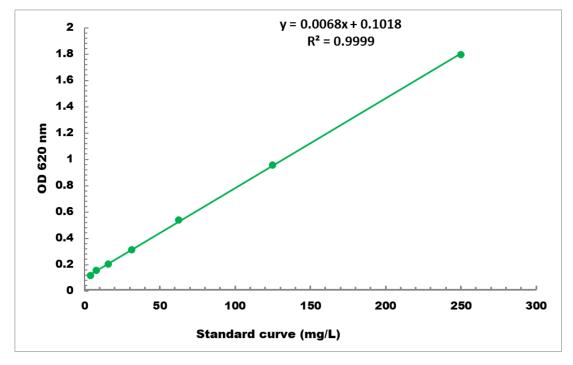
V_{Standard}: the volume of standard, 0.01 ml;

 V_{Sample} : the volume of sample, 0.01 ml.



VII. TYPICAL DATA

The standard curve is for demonstration only. A standard curve must be run with each assay.



Detection Range: 5 mg/L-250 mg/L

VIII. TECHNICAL SUPPORT

For troubleshooting, information or assistance, please go online towww.cohesionbio.com or contact us at techsupport@cohesionbio.com

IX. NOTES