



# **Proline Microplate Assay Kit**

## **User Manual**

**Catalog # CAK1121**

(Version 1.2A)

Detection and Quantification of Proline content in Urine, Serum, Plasma, Tissue extracts, Cell lysate, Cell culture media and Other biological fluids Samples.

**For research use only. Not for diagnostic or therapeutic procedures.**

I. INTRODUCTION.....2

II. KIT COMPONENTS.....3

III. MATERIALS REQUIRED BUT NOT PROVIDED.....3

IV. SAMPLE PREPARATION.....4

V. ASSAY PROCEDURE.....5

VI. CALCULATION.....6

VII. TYPICAL DATA.....7

VIII. TECHNICAL SUPPORT.....7

IX. NOTES.....7

## I. INTRODUCTION

Proline (abbreviated as Pro or P; encoded by the codons CCU, CCC, CCA, and CCG) is an  $\alpha$ -amino acid that is used in the biosynthesis of proteins. It contains an  $\alpha$ -amino group (which is in the protonated  $>NH_2^+$  form under biological conditions), an  $\alpha$ -carboxylic acid group (which is in the deprotonated  $-COO^-$  form under biological conditions), and a side chain pyrrolidine, classifying it as a nonpolar (at physiological pH), aliphatic amino acid. It is non-essential in humans, meaning the body can synthesize it from the non-essential amino acid L-glutamate.

Proline Microplate Assay Kit is a sensitive assay for determining Proline in various samples. Proline concentration is determined by Ninhydrin. The reaction products can be measured at a colorimetric readout at 520 nm.

## II. KIT COMPONENTS

Component	Volume	Storage
96-Well Microplate	1 plate	
Assay Buffer	30 ml x 4	4 °C
Reaction Buffer	5 ml x 1	4 °C
Dye Reagent	Powder x 1	4 °C
Dye Reagent Diluent	5 ml x 1	4 °C
Standard	Powder x 1	4 °C
Plate Adhesive Strips	3 Strips	
Technical Manual	1 Manual	

### Note:

**Dye Reagent:** Add 5 ml Dye Reagent Diluent into Dye Reagent bottle, heat to dissolve.

**Standard:** Add 1 ml distilled water to dissolve before use, then add 0.1 ml into 0.9 ml distilled water, the concentration will be 200 µg/ml.

## III. MATERIALS REQUIRED BUT NOT PROVIDED

1. Microplate reader to read absorbance at 520 nm
2. Distilled water
3. Pipettor, multi-channel pipettor
4. Pipette tips
5. Mortar
6. Centrifuge
7. Timer
8. Convection oven

#### IV. SAMPLE PREPARATION

##### 1. For cell and bacteria samples

Collect cell or bacteria into centrifuge tube, discard the supernatant after centrifugation, add 1 ml Assay buffer for  $5 \times 10^6$  cell or bacteria, sonicate (with power 20%, sonicate 3s, interval 10s, repeat 30 times); put it into boiling water bath for 10 minutes; centrifuged at 10000g for 10 minutes, take the supernatant into a new centrifuge tube for detection.

##### 2. For tissue samples

Weigh out 0.1 g tissue, homogenize with 1 ml Assay buffer; then transfer into centrifuge tube, put it into boiling water bath for 10 minutes; centrifuged at 10000g 4 °C for 10 minutes, take the supernatant into a new centrifuge tube for detection.

##### 3. For serum or plasma samples

Add 0.1 ml serum or plasma and 0.9 ml Assay buffer into the microcentrifuge tube, mix; put it into boiling water bath for 10 minutes; centrifuged at 10000g for 10 minutes, take the supernatant into a new centrifuge tube for detection.

## V. ASSAY PROCEDURE

Add following reagents into the microplate:

Reagent	Sample	Standard	Blank
Sample	50 $\mu$ l	--	--
Standard	--	50 $\mu$ l	--
Distilled water	--	--	50 $\mu$ l
Reaction Buffer	50 $\mu$ l	50 $\mu$ l	50 $\mu$ l
Dye Reagent	50 $\mu$ l	50 $\mu$ l	50 $\mu$ l
Mix, put it into the convection oven, 90 °C for 20 minutes, mix, record absorbance measured at 520 nm.			

### Note:

- 1) Perform 2-fold serial dilutions of the top standards to make the standard curve.
- 2) The concentrations can vary over a wide range depending on the different samples. For unknown samples, we recommend doing a pilot experiment & testing several doses to ensure the readings are within the standard curve range.
- 3) Reagents must be added step by step, can not be mixed and added together.

## VI. CALCULATION

1. According to the protein concentration of sample

$$\begin{aligned} \text{Pro } (\mu\text{g}/\text{mg}) &= (C_{\text{Standard}} \times V_{\text{Standard}}) \times (\text{OD}_{\text{Sample}} - \text{OD}_{\text{Blank}}) / (\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}}) / (V_{\text{Sample}} \\ &\quad \times C_{\text{Protein}}) \\ &= 200 \times (\text{OD}_{\text{Sample}} - \text{OD}_{\text{Blank}}) / (\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}}) / C_{\text{Protein}} \end{aligned}$$

2. According to the weight of sample

$$\begin{aligned} \text{Pro } (\mu\text{g}/\text{g}) &= (C_{\text{Standard}} \times V_{\text{Standard}}) \times (\text{OD}_{\text{Sample}} - \text{OD}_{\text{Blank}}) / (\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}}) / (W \times \\ &\quad V_{\text{Sample}} / V_{\text{Total}}) \\ &= 200 \times (\text{OD}_{\text{Sample}} - \text{OD}_{\text{Blank}}) / (\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}}) / W \end{aligned}$$

3. According to the quantity of cells or bacteria

$$\begin{aligned} \text{Pro } (\mu\text{g}/10^4) &= (C_{\text{Standard}} \times V_{\text{Standard}}) \times (\text{OD}_{\text{Sample}} - \text{OD}_{\text{Blank}}) / (\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}}) / (V_{\text{Sample}} \\ &\quad \times N / V_{\text{Assay}}) \\ &= 200 \times (\text{OD}_{\text{Sample}} - \text{OD}_{\text{Blank}}) / (\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}}) / N \end{aligned}$$

4. According to the volume of serum or plasma

$$\begin{aligned} \text{Pro } (\mu\text{g}/\text{ml}) &= (C_{\text{Standard}} \times V_{\text{Standard}}) \times (\text{OD}_{\text{Sample}} - \text{OD}_{\text{Blank}}) / (\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}}) / (V_{\text{Sample}} \\ &\quad \times V / V_{\text{Assay}}) \\ &= 200 \times (\text{OD}_{\text{Sample}} - \text{OD}_{\text{Blank}}) / (\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}}) / V \end{aligned}$$

$C_{\text{Standard}}$ : the standard concentration, 200  $\mu\text{g}/\text{ml}$ ;

$C_{\text{Protein}}$ : the protein concentration,  $\text{mg}/\text{ml}$ ;

W: the weight of sample, g;

V: the volume of serum or plasma, ml;

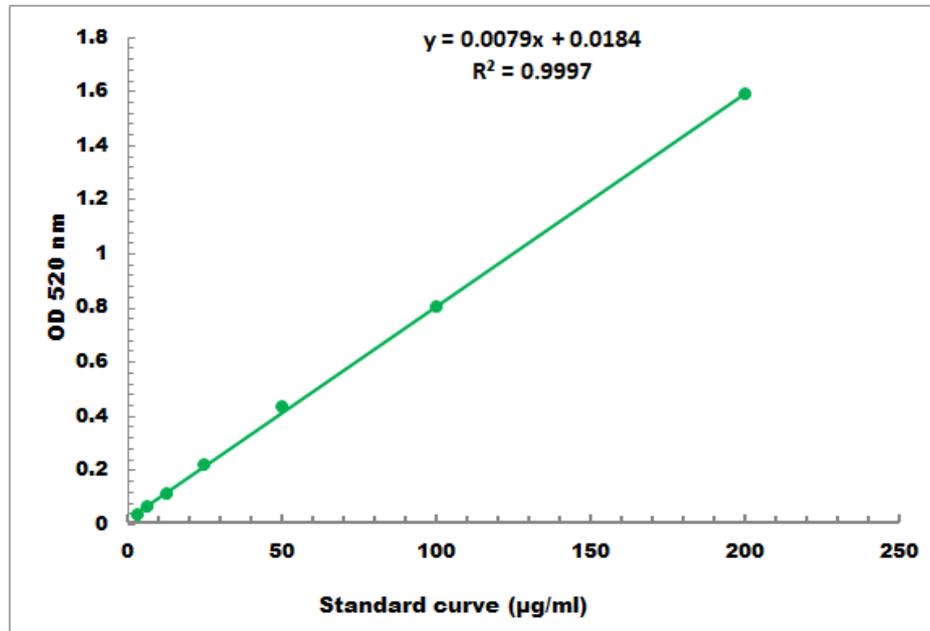
$V_{\text{Standard}}$ : the volume of standard, 0.05 ml;

$V_{\text{Sample}}$ : the volume of sample, 0.05 ml;

$V_{\text{Assay}}$ : the volume of Assay buffer, 1 ml.

## VII. TYPICAL DATA

The standard curve is for demonstration only. A standard curve must be run with each assay.



Detection Range: 2 µg/ml - 200 µg/ml

## VIII. TECHNICAL SUPPORT

For troubleshooting, information or assistance, please go online to [www.cohesionbio.com](http://www.cohesionbio.com) or contact us at [techsupport@cohesionbio.com](mailto:techsupport@cohesionbio.com)

## IX. NOTES