



Iron Microplate Assay Kit

User Manual

Catalog # CAK1106

(Version 1.6D)

Detection and Quantification of Iron (Fe^{3+}) Content in Serum, Plasma, Urine, Saliva, Tissue extracts, Cell lysate and Other biological fluids Samples.

For research use only. Not for diagnostic or therapeutic procedures.

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I. INTRODUCTION

Iron level in blood is a reliable diagnostic indicator of various disease states.

Increased levels of iron concentration in blood are associated with blood loss, increased destruction of red blood cells (e.g. hemorrhage) or decreased blood cell survival, acute hepatitis, certain sideroachrestic anemias, ingestion of iron-rich diets, defects in iron storage (e.g. pernicious anemia). Decreased levels of blood iron may result from insufficient iron ingestion from diets, chronic blood loss pathologies, or increased demand on iron storage as during normal pregnancy.

Iron Microplate Assay Kit provides a simple and direct procedure for measuring iron (I) levels in a variety of samples. The ferrum ions can react with Phenanthroline. The products can be measured at a colorimetric readout at 510 nm.

II. KIT COMPONENTS

Component	Volume	Storage
96-Well Microplate	1 plate	
Assay Buffer	30 ml x 4	4 °C
Reducing Reagent	Powder x 1	4 °C
Reaction Buffer	5 ml x 1	4 °C
Dye Reagent	Powder x 1	4 °C
Standard (500 µmol/L)	1 ml x 1	4 °C
Plate Adhesive Strips	3 Strips	
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Note:

Reducing Reagent: add 2.5 ml distilled water to dissolve before use.

Dye Reagent: add 2.5 ml distilled water to dissolve before use.

III. MATERIALS REQUIRED BUT NOT PROVIDED

1. Microplate reader to read absorbance at 510 nm
2. Distilled water
3. Pipettor, multi-channel pipettor
4. Pipette tips
5. Centrifuge
6. Timer

IV. SAMPLE PREPARATION

1. For liquid sample

Liquid samples can be tested directly.

Serum or plasma samples, add 0.5 ml Assay Buffer into 0.5 ml sample, mix, centrifuged at 10000g for 10 minutes, take the supernatant into a new centrifuge tube for detection.

2. For cell and bacteria samples

Collect cell or bacteria into centrifuge tube, discard the supernatant after centrifugation, add 0.5 ml ddH₂O for 5×10^6 cell or bacteria, sonicate (with power 20%, sonicate 3s, interval 10s, repeat 30 times); then add 0.5 ml Assay Buffer mix, centrifuged at 10000g for 10 minutes, take the supernatant into a new centrifuge tube for detection.

3. For tissue samples

Weigh out 0.1 g tissue, homogenize with 0.5 ml ddH₂O, then add 0.5 ml Assay Buffer mix, centrifuged at 10000g for 10 minutes, take the supernatant into a new centrifuge tube for detection.

V. ASSAY PROCEDURE

Warm all the reagents to room temperature before use.

Add following reagents into the microplate:

Reagent	Sample	Standard	Blank
Sample	100 μ l	--	--
Standard	--	100 μ l	--
Distilled water	--	--	100 μ l
Reducing Reagent	25 μ l	25 μ l	25 μ l
Reaction Buffer	50 μ l	50 μ l	50 μ l
Dye Reagent	25 μ l	25 μ l	25 μ l
Mix, incubate at 37 °C for 60 mins, measured at 510 nm and record the absorbance.			

Note:

- 1) Perform 2-fold serial dilutions of the top standards to make the standard curve.
- 2) The concentrations can vary over a wide range depending on the different samples.
For unknown samples, we recommend doing a pilot experiment & testing several doses to ensure the readings are within the standard curve range.
- 3) Reagents must be added step by step, can not be mixed and added together.

VI. CALCULATION

1. According to the volume of sample

$$\begin{aligned} \text{Iron } (\mu\text{mol/ml}) &= (C_{\text{Standard}} \times V_{\text{Standard}}) \times (OD_{\text{Sample}} - OD_{\text{Blank}}) / (OD_{\text{Standard}} - OD_{\text{Blank}}) / \\ &V_{\text{Sample}} \times 2 \\ &= (OD_{\text{Sample}} - OD_{\text{Blank}}) / (OD_{\text{Standard}} - OD_{\text{Blank}}) \end{aligned}$$

2. According to the weight of sample

$$\begin{aligned} \text{Iron } (\mu\text{mol/g}) &= (C_{\text{Standard}} \times V_{\text{Standard}}) \times (OD_{\text{Sample}} - OD_{\text{Blank}}) / (OD_{\text{Standard}} - OD_{\text{Blank}}) / \\ &(V_{\text{Sample}} \times W / V_{\text{Assay}}) \\ &= 0.5 \times (OD_{\text{Sample}} - OD_{\text{Blank}}) / (OD_{\text{Standard}} - OD_{\text{Blank}}) / W \end{aligned}$$

3. According to the quantity of cells or bacteria

$$\begin{aligned} \text{Iron } (\mu\text{mol}/10^4) &= (C_{\text{Standard}} \times V_{\text{Standard}}) \times (OD_{\text{Sample}} - OD_{\text{Blank}}) / (OD_{\text{Standard}} - OD_{\text{Blank}}) / (N \times \\ &V_{\text{Sample}} / V_{\text{Assay}}) \\ &= 0.5 \times (OD_{\text{Sample}} - OD_{\text{Blank}}) / (OD_{\text{Standard}} - OD_{\text{Blank}}) / N \end{aligned}$$

C_{Standard} : the concentration of Standard, 500 $\mu\text{mol/L}$ = 0.5 $\mu\text{mol/ml}$;

W : the weight of sample, g;

N : the quantity of cell or bacteria, $N \times 10^4$;

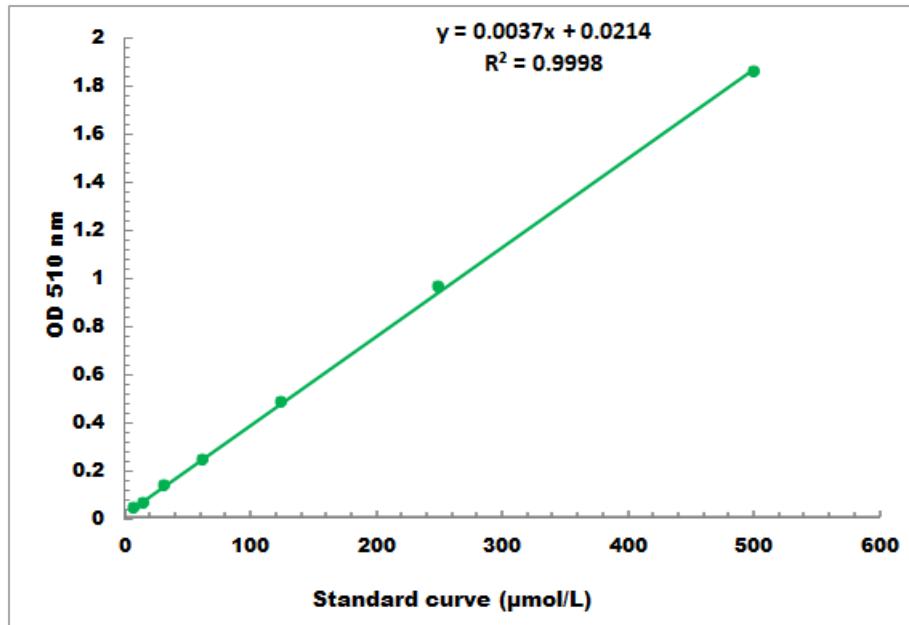
V_{Standard} : the volume of standard, 0.1 ml;

V_{Sample} : the volume of sample, 0.1 ml;

V_{Assay} : the volume of ddH₂O + Assay Buffer, 1 ml.

VII. TYPICAL DATA

The standard curve is for demonstration only. A standard curve must be run with each assay.



Detection Range: 5 µmol/L - 500 µmol/L

VIII. TECHNICAL SUPPORT

For troubleshooting, information or assistance, please go online to www.cohesionbio.com or contact us at techsupport@cohesionbio.com

IX. NOTES