

Pyruvate Decarboxylase Microplate Assay Kit

User Manual

Catalog # CAK1085

(Version 1.4D)

Detection and Quantification of Pyruvate Decarboxylase (PDC) Activity in Urine, Serum, Plasma, Tissue extracts, Cell lysate, Cell culture media and Other biological fluids Samples.

For research use only. Not for diagnostic or therapeutic procedures.



I. INTRODUCTION	2
II. KIT COMPONENTS	3
III. MATERIALS REQUIRED BUT NOT PROVIDED	3
IV. SAMPLE PREPARATION	4
V. ASSAY PROCEDURE	5
VI. CALCULATION	6
VII. TYPICAL DATA	7
VIII. TECHNICAL SUPPORT	7
IX. NOTES	7



I. INTRODUCTION

Pyruvate decarboxylase catalyzes the oxidation of decarboxylation of pyruvate to acetaldehyde. This assay is an indirect method in which the conversion is linked to the activity of the subsequent enzyme alcohol dehydrogenase, which supplied in excess, converts the product acetaldehyde effectively into NAD and ethanol. Pyruvate Decarboxylase Microplate Assay Kit provides a simple and direct procedure for measuring pyruvate decarboxylase activity in a variety of samples. The reaction velocity is determined by the rate of absorbance at 340 nm resulting from reduction of NADH is measured.



II. KIT COMPONENTS

Component	Volume	Storage
96-Well Microplate	1 plate	
Assay Buffer	30 ml x 4	4 °C
Reaction Buffer	20 ml x 1	4 °C
Substrate	Powder x 1	4 °C
Enzyme	Powder x 1	-20 °C
Standard	Powder x 1	-20 °C
Positive Control	5 μl x 1	4 °C
Technical Manual	1 Manual	

Note:

Substrate: add 1 ml Reaction Buffer to dissolve before use.

Enzyme: add 1 ml Reaction Buffer to dissolve before use.

Standard: add 1 ml distilled water to dissolve before use; then add 0.2 ml into 0.8 ml

distilled water, the concentration will be 400 μ mol/L.

Positive Control: add 50 µl Assay Buffer before use.

III. MATERIALS REQUIRED BUT NOT PROVIDED

- 1. Microplate reader to read absorbance at 340 nm
- 2. Distilled water
- 3. Pipettor, multi-channel pipettor
- 4. Pipette tips
- 5. Mortar
- 6. Centrifuge
- 7. Timer
- 8. Ice



IV. SAMPLE PREPARATION

1. For cell and bacteria samples

Collect cell or bacteria into centrifuge tube, discard the supernatant after centrifugation, add 1 ml Assay buffer for 5×10^6 cell or bacteria, sonicate (with power 20%, sonicate 3s, interval 10s, repeat 30 times); centrifuged at 16,000g 4 °C for 20 minutes, take the supernatant into a new centrifuge tube and keep it on ice for detection.

2. For tissue samples

Weigh out 0.1 g tissue, homogenize with 1 ml Assay buffer on ice, centrifuged at 16,000g 4 °C for 20 minutes, take the supernatant into a new centrifuge tube and keep it on ice for detection.

For serum or plasma samples
Detect directly.



V. ASSAY PROCEDURE

Warm all reagents to room temperature before use.

Add following reagents into the microplate:

Reagent	Sample	Standard	Blank	Positive		
				Control		
Standard		200 µl				
Distilled water			200 µl			
Reaction Buffer	170 µl			170 µl		
Substrate	10 µl			10 µl		
Enzyme	10 µl			10 µl		
Mix.						
Sample	10 µl					
Positive Control				10 µl		
Mix, measured at 340 nm and record the absorbance of 10th second and 130th						
second.						

Note:

1) Perform 2-fold serial dilutions of the top standards to make the standard curve.

2) For unknown samples, we recommend doing a pilot experiment & testing several doses to ensure the readings are within the standard curve range. If the enzyme activity is lower, please add more sample into the reaction system; or increase the reaction time; if the enzyme activity is higher, please dilute the sample, or decrease the reaction time.

3) Reagents must be added step by step, can not be mixed and added together.



VI. CALCULATION

Unit Definition: One unit of PDC activity is defined as the enzyme that decomposes 1 μ mol of NADH per minute.

1. According to the protein concentration of sample

 $PDC (U/mg) = (C_{Standard} \times V_{Standard}) \times (OD_{Sample(10S)} - OD_{Sample(130S)}) / (OD_{Standard} - OD_{Blank}) / (V_{Sample} \times C_{Protein}) / T$

= $4 \times (OD_{Sample(10S)} - OD_{Sample(130S)}) / (OD_{Standard} - OD_{Blank}) / C_{Protein}$

2. According to the weight of sample

 $PDC (U/g) = (C_{Standard} \times V_{Standard}) \times (OD_{Sample(10S)} - OD_{Sample(130S)}) / (OD_{Standard} - OD_{Blank}) / (OD_{Standard} - OD_{Standard} -$

(V_{Sample} × W / V_{Assay}) / T

= $4 \times (OD_{Sample(10S)} - OD_{Sample(130S)}) / (OD_{Standard} - OD_{Blank}) / W$

3. According to the quantity of cells or bacteria

 $PDC (U/10^{4}) = (C_{Standard} \times V_{Standard}) \times (OD_{Sample(10S)} - OD_{Sample(13OS)}) / (OD_{Standard} - OD_{Blank})$

/ (V_{Sample} × N / V_{Assay}) / T

= $4 \times (OD_{Sample(10S)} - OD_{Sample(130S)}) / (OD_{Standard} - OD_{Blank}) / N$

4. According to the volume of serum or plasma

PDC (U/ml) = (C_{Standard} × V_{Standard}) × (OD_{Sample(10S)} - OD_{Sample(130S)}) / (OD_{Standard} - OD_{Blank}) /

V_{Sample} / T

= $4 \times (OD_{Sample(10S)} - OD_{Sample(130S)}) / (OD_{Standard} - OD_{Blank})$

 C_{Standard} : the standard concentration, 400 µmol/L = 0.4 µmol/ml;

 V_{Standard} : the volume of standard, 200 µl = 0.2 ml;

C_{Protein}: the protein concentration, mg/ml;

W: the weight of sample, g;

N: the quantity of cell or bacteria, $N \times 10^4$;

V_{Sample}: the volume of sample, 0.01 ml;

V_{Assay}: the volume of Assay buffer, 1 ml;

T: the reaction time, 2 minutes.



VII. TYPICAL DATA

The standard curve is for demonstration only. A standard curve must be run with each assay.



Detection Range: 4 µmol/L - 400 µmol/L



Positive Control reaction in 96-well plate assay with decreasing the concentration

VIII. TECHNICAL SUPPORT

For troubleshooting, information or assistance, please go online to www.cohesionbio.com or contact us at techsupport@cohesionbio.com

IX. NOTES