



# **Ascorbate Peroxidase Microplate Assay Kit User Manual**

**Catalog # CAK1052**

(Version 1.1C)

Detection and Quantification of Ascorbate Peroxidase (APX) Activity  
in Tissue extracts, Cell lysate Samples.

**For research use only. Not for diagnostic or therapeutic procedures.**

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## I. INTRODUCTION

Ascorbate peroxidase is a hydrogen peroxide-scavenging enzyme that is specific to plants and algae and is indispensable to protect chloroplasts and other cell constituents from damage by hydrogen peroxide and hydroxyl radicals produced from it.

The assay is initiated with the enzymatic oxidation of AsA by APX. AsA can be measured at a colorimetric readout at 290 nm.

## II. KIT COMPONENTS

Component	Volume	Storage
96-Well UV Microplate	1 plate	
Assay Buffer	30 ml x 4	4 °C
Substrate I	Powder x 1	4 °C
Substrate II	5 ml x 1	4 °C
Technical Manual	1 Manual	

**Note:**

**Substrate I:** add 13 ml Assay Buffer to dissolve before use.

## III. MATERIALS REQUIRED BUT NOT PROVIDED

1. Microplate reader to read absorbance at 290 nm
2. Distilled water
3. Pipettor, multi-channel pipettor
4. Pipette tips
5. Mortar
6. Centrifuge
7. Timer
8. Ice

#### IV. SAMPLE PREPARATION

##### 1. For cell and bacteria samples

Collect cell or bacteria into centrifuge tube, discard the supernatant after centrifugation, add 1 ml Assay buffer for  $5 \times 10^6$  cell or bacteria, sonicate (with power 20%, sonicate 3s, interval 10s, repeat 30 times); centrifuged at 13000g 4 °C for 20 minutes, take the supernatant into a new centrifuge tube and keep it on ice for detection.

##### 2. For tissue samples

Weigh out 0.1 g tissue, homogenize with 1 ml Assay buffer on ice, centrifuged at 13000g 4 °C for 20 minutes, take the supernatant into a new centrifuge tube and keep it on ice for detection.

## V. ASSAY PROCEDURE

Warm the Substrate I to room temperature before use.

Add following reagents into the microplate:

Reagent	Sample	Blank
Sample	20 $\mu$ l	--
Distilled water	--	20 $\mu$ l
Substrate I	130 $\mu$ l	130 $\mu$ l
Substrate II	50 $\mu$ l	50 $\mu$ l
Mix, measured at 290 nm and record the absorbance of 10th second and 310th second.		

1) Reagents must be added step by step, can not be mixed and added together.

## VI. CALCULATION

**Unit Definition:** One unit of APX is the amount of enzyme that will oxidize 1  $\mu\text{mol}$  AsA per minute.

1. According to the protein concentration of sample

$$\begin{aligned} \text{APX (U/mg)} &= \frac{[(\text{OD}_{\text{Sample}(10\text{S})} - \text{OD}_{\text{Sample}(310\text{S})}) - (\text{OD}_{\text{Blank}(10\text{S})} - \text{OD}_{\text{Blank}(310\text{S})})]}{(\epsilon \times d) \times V_{\text{Total}}} \\ &\quad / (V_{\text{Sample}} \times C_{\text{Protein}}) / T \\ &= 1.19 \times \frac{[(\text{OD}_{\text{Sample}(10\text{S})} - \text{OD}_{\text{Sample}(310\text{S})}) - (\text{OD}_{\text{Blank}(10\text{S})} - \text{OD}_{\text{Blank}(310\text{S})})]}{C_{\text{Protein}}} \end{aligned}$$

2. According to the weight of sample

$$\begin{aligned} \text{APX (U/g)} &= \frac{[(\text{OD}_{\text{Sample}(10\text{S})} - \text{OD}_{\text{Sample}(310\text{S})}) - (\text{OD}_{\text{Blank}(10\text{S})} - \text{OD}_{\text{Blank}(310\text{S})})]}{(\epsilon \times d) \times V_{\text{Total}}} / \\ &\quad (W \times V_{\text{Sample}} / V_{\text{Assay}}) / T \\ &= 1.19 \times \frac{[(\text{OD}_{\text{Sample}(10\text{S})} - \text{OD}_{\text{Sample}(310\text{S})}) - (\text{OD}_{\text{Blank}(10\text{S})} - \text{OD}_{\text{Blank}(310\text{S})})]}{W} \end{aligned}$$

3. According to the quantity of cells or bacteria

$$\begin{aligned} \text{APX (U}/10^4) &= \frac{[(\text{OD}_{\text{Sample}(10\text{S})} - \text{OD}_{\text{Sample}(310\text{S})}) - (\text{OD}_{\text{Blank}(10\text{S})} - \text{OD}_{\text{Blank}(310\text{S})})]}{(\epsilon \times d) \times V_{\text{Total}}} \\ &\quad / (N \times V_{\text{Sample}} / V_{\text{Assay}}) / T \\ &= 1.19 \times \frac{[(\text{OD}_{\text{Sample}(10\text{S})} - \text{OD}_{\text{Sample}(310\text{S})}) - (\text{OD}_{\text{Blank}(10\text{S})} - \text{OD}_{\text{Blank}(310\text{S})})]}{N} \end{aligned}$$

$\epsilon$ : molar extinction coefficient,  $2.8 \times 10^3 \text{ L/mol/cm} = 2.8 \text{ ml}/\mu\text{mol/cm}$ ;

$d$ : the optical path of 96-Well microplate, 0.6 cm;

$C_{\text{Protein}}$ : the protein concentration, mg/ml;

$W$ : the weight of sample, g;

$N$ : the quantity of cell or bacteria,  $N \times 10^4$ ;

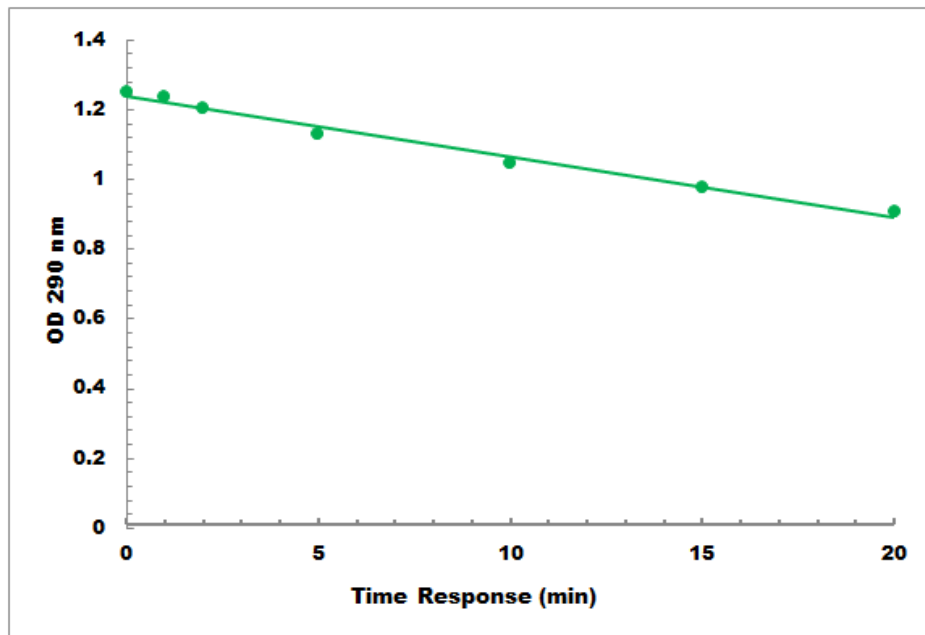
$V_{\text{Total}}$ : the total volume of the enzymatic reaction, 0.2 ml;

$V_{\text{Sample}}$ : the volume of sample, 0.02 ml;

$V_{\text{Assay}}$ : the volume of Assay buffer, 1 ml;

$T$ : the reaction time, 5 minutes.

## VII. TYPICAL DATA



Samples were assayed using the 96-well microplate

## VIII. TECHNICAL SUPPORT

For troubleshooting, information or assistance, please go online to [www.cohesionbio.com](http://www.cohesionbio.com) or contact us at [techsupport@cohesionbio.com](mailto:techsupport@cohesionbio.com)

## IX. NOTES