

Glutathione Reductase Microplate Assay Kit User Manual

Catalog # CAK1043

(Version 1.3D)

Detection and Quantification of Glutathione Reductase (GR) Activity in Urine, Serum, Plasma, Other biological fluids, Tissue extracts, Cell lysate, Cell culture media Samples.

For research use only. Not for diagnostic or therapeutic procedures.



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I. INTRODUCTION

Glutathione reductase (GR, EC 1.6.4.2) is a flavoprotein that catalyzes the NADPH-dependent reduction of oxidized glutathione (GSSG) to glutathione (GSH). This enzyme is essential for the GSH redox cycle which maintains adequate levels of reduced cellular GSH. A high GSH/GSSG ratio is essential for protection against oxidative stress.

Glutathione Reductase Microplate Assay Kit measures GR activity by measuring the rate of NADPH oxidation. The oxidation of NADPH to NADP+ is accompanied by a decrease in absorbance at 340 nm. Since GR is present at rate limiting concentrations, the rate of decrease in the A340 is directly proportional to the GR activity in the sample.



II. KIT COMPONENTS

Component	Volume	Storage
96-Well Microplate	1 plate	
Assay Buffer	30 ml x 4	4 °C
Substrate	Powder x 1	-20 °C
Standard	Powder x 1	-20 °C
Positive Control	10 μΙ	4 °C
Technical Manual	1 Manual	

Note:

Substrate: add 19 ml Assay Buffer to dissolve before use.

Standard: add 1 ml distilled water to dissolve before use; then add 0.2 ml into 0.8 ml

distilled water, the concentration will be 400 μ mol/L.

Positive Control: add 90 μ l distilled water to dissolve before use.

III. MATERIALS REQUIRED BUT NOT PROVIDED

- 1. Microplate reader to read absorbance at 340 nm
- 2. Distilled water
- 3. Pipettor, multi-channel pipettor
- 4. Pipette tips
- 5. Mortar
- 6. Centrifuge
- 7. Timer
- 8. Ice



IV. SAMPLE PREPARATION

1. For cell and bacteria samples

Collect cell or bacteria into centrifuge tube, discard the supernatant after centrifugation, add 1 ml Assay buffer for 5×10^6 cell or bacteria, sonicate (with power 20%, sonicate 3s, interval 10s, repeat 30 times); centrifuged at 8,000g 4 °C for 10 minutes, take the supernatant into a new centrifuge tube and keep it on ice for detection.

2. For tissue samples

Weigh out 0.1 g tissue, homogenize with 1 ml Assay buffer on ice, centrifuged at 8,000g 4 °C for 20 minutes, take the supernatant into a new centrifuge tube and keep it on ice for detection.

3. For serum or plasma samples

Detect directly.



V. ASSAY PROCEDURE

Warm all reagents to room temperature before use.

Add following reagents in the microplate:

Reagent	Sample	Standard	Blank	Positive
				Control
Standard		200 μΙ		
Distilled water			200 μΙ	
Substrate	190 μΙ			190 μΙ
Sample	10 μΙ			
Positive Control				10 μΙ

Mix, measured at 340 nm and record the absorbance of 10th second and 130th second.

Note:

- 1) Perform 2-fold serial dilutions of the top standards to make the standard curve.
- 2) For unknown samples, we recommend doing a pilot experiment & testing several doses to ensure the readings are within the standard curve range. If the enzyme activity is lower, please add more sample into the reaction system; or increase the reaction time; if the enzyme activity is higher, please dilute the sample, or decrease the reaction time.
- 3) Reagents must be added step by step, can not be mixed and added together.



VI. CALCULATION

Unit Definition: One unit of GR activity is defined as the enzyme that reduces 1 μ mol of NADPH per minute.

1. According to the protein concentration of sample

$$\begin{aligned} \mathsf{GR} \; (\mathsf{U/mg}) &= \left(\mathsf{C}_{\mathsf{Standard}} \times \mathsf{V}_{\mathsf{Standard}} \right) \times \left(\mathsf{OD}_{\mathsf{Sample}(10S)} - \mathsf{OD}_{\mathsf{Sample}(130S)} \right) / \left(\mathsf{OD}_{\mathsf{Standard}} - \mathsf{OD}_{\mathsf{Blank}} \right) / \\ &= 4 \times \left(\mathsf{OD}_{\mathsf{Sample}(10S)} - \mathsf{OD}_{\mathsf{Sample}(130S)} \right) / \left(\mathsf{OD}_{\mathsf{Standard}} - \mathsf{OD}_{\mathsf{Blank}} \right) / \mathsf{C}_{\mathsf{Protein}} \end{aligned}$$

2. According to the weight of sample

$$\begin{split} \text{GR (U/g) = (C}_{\text{Standard}} \times \text{V}_{\text{Standard}}) \times & \left(\text{OD}_{\text{Sample(10S)}} - \text{OD}_{\text{Sample(130S)}} \right) / \left(\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}} \right) / \\ & \left(\text{V}_{\text{Sample}} \times \text{W} / \text{V}_{\text{Assay}} \right) / \text{T} \\ & = 4 \times \left(\text{OD}_{\text{Sample(10S)}} - \text{OD}_{\text{Sample(130S)}} \right) / \left(\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}} \right) / \text{W} \end{split}$$

3. According to the quantity of cells or bacteria

$$\begin{aligned} \mathsf{GR} \; (\mathsf{U}/10^4) &= \left(\mathsf{C}_{\mathsf{Standard}} \times \mathsf{V}_{\mathsf{Standard}}\right) \times \left(\mathsf{OD}_{\mathsf{Sample}(10\mathsf{S})} - \mathsf{OD}_{\mathsf{Sample}(130\mathsf{S})}\right) / \left(\mathsf{OD}_{\mathsf{Standard}} - \mathsf{OD}_{\mathsf{Blank}}\right) / \\ & \left(\mathsf{V}_{\mathsf{Sample}} \times \mathsf{N} \; / \; \mathsf{V}_{\mathsf{Assay}}\right) / \; \mathsf{T} \\ &= 4 \times \left(\mathsf{OD}_{\mathsf{Sample}(10\mathsf{S})} - \mathsf{OD}_{\mathsf{Sample}(130\mathsf{S})}\right) / \left(\mathsf{OD}_{\mathsf{Standard}} - \mathsf{OD}_{\mathsf{Blank}}\right) / \; \mathsf{N} \end{aligned}$$

4. According to the volume of sample

GR (U/mI) = (
$$C_{Standard} \times V_{Standard}$$
) × ($OD_{Sample(10S)}$ - $OD_{Sample(130S)}$) / ($OD_{Standard}$ - OD_{Blank}) / V_{Sample} / T = 4 × ($OD_{Sample(10S)}$ - $OD_{Sample(130S)}$) / ($OD_{Standard}$ - OD_{Blank})

 $C_{Standard}$: the standard concentration, 400 μ mol/L = 0.4 μ mol/ml;

 $V_{Standard}$: the volume of standard, 200 μ l = 0.2 ml;

C_{Protein}: the protein concentration, mg/ml;

W: the weight of sample, g;

N: the quantity of cell or bacteria, $N \times 10^4$;

V_{Sample}: the volume of sample, 0.01 ml;

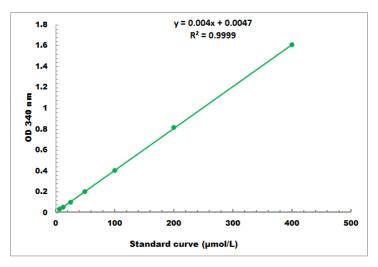
V_{Assay}: the volume of Assay buffer, 1 ml;

T: the reaction time, 2 minutes.

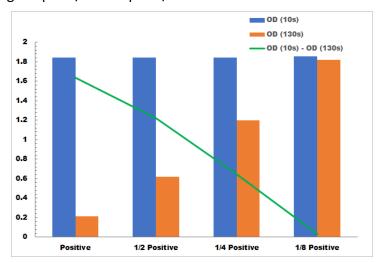


VII. TYPICAL DATA

The standard curve is for demonstration only. A standard curve must be run with each assay.



Detection Range: 4 μmol/L - 400 μmol/L



Positive Control reaction in 96-well plate assay with decreasing the concentration

VIII. TECHNICAL SUPPORT

For troubleshooting, information or assistance, please go online to www.cohesionbio.com or contact us at techsupport@cohesionbio.com

IX. NOTE