Detection and Quantification of Nitrate Reductase (NR) Activity in Tissue extracts, Cell lysate Samples.

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I. INTRODUCTION

Nitrate Reductase (NR), also known as respiratory nitrate reductase. Eukaryotic nitrate reductases are part of the sulfite oxidase family of molybdoenzymes. They transfer electrons from NADH or NADPH to nitrate. Prokaryotic nitrate reductases belong to the DMSO reductase family of molybdoenzymes and have been classified into three groups, assimilatory nitrate reductases (Nas), respiratory nitrate reductase (Nar), and periplasmic nitrate reductases (Nap). The active site of these enzymes is a Mo ion that is bound to the four thiolate functions of two pterin molecules. The coordination sphere of the Mo is completed by one amino-acid side chain and oxygen and/or sulfur ligands. The exact environment of the Mo ion in certain of these enzymes (oxygen versus sulfur as a sixth molybdenum ligand) is still debated. The Mo is covalently attached to the protein by a cysteine ligand in Nap, and an aspartate in Nar. Nitrate reductase activity can be used as a biochemical tool for predicting grain yield and grain protein production. Nitrate reductase promotes amino acid production in tea leaves. It is reported that tea plants sprayed with various micronutrients (like Zn, Mn and B) along with Mo enhanced the amino acid content of tea shoots and also the crop yield.

The assay is initiated with the enzymatic hydrolysis of the nitrate by Nitrate Reductase. The enzyme catalysed reaction products azo-compound can be measured at a colorimetric readout at 540 nm.
II. KIT COMPONENTS

<table>
<thead>
<tr>
<th>Component</th>
<th>Volume</th>
<th>Storage</th>
</tr>
</thead>
<tbody>
<tr>
<td>96-Well Microplate</td>
<td>1 plate</td>
<td></td>
</tr>
<tr>
<td>Assay Buffer</td>
<td>30 ml x 4</td>
<td>4 °C</td>
</tr>
<tr>
<td>Substrate I</td>
<td>6 ml x 1</td>
<td>4 °C</td>
</tr>
<tr>
<td>Substrate II</td>
<td>2 ml x 1</td>
<td>-20 °C</td>
</tr>
<tr>
<td>Dye Reagent I</td>
<td>5 ml x 1</td>
<td>4 °C</td>
</tr>
<tr>
<td>Dye Reagent II</td>
<td>5 ml x 1</td>
<td>4 °C</td>
</tr>
<tr>
<td>Standard (10 μmol/ml)</td>
<td>1 ml x 1</td>
<td>4 °C</td>
</tr>
<tr>
<td>Plate Adhesive Strips</td>
<td>3 Strips</td>
<td></td>
</tr>
</tbody>
</table>

III. MATERIALS REQUIRED BUT NOT PROVIDED

1. Microplate reader to read absorbance at 540 nm
2. Distilled water
3. Pipettor
4. Pipette tips
5. Mortar
6. Ice
7. Centrifuge
8. Timer
IV. SAMPLE PREPARATION

1. For tissue samples
Weigh 0.1 g tissue, homogenize with 1 ml Assay buffer on ice, centrifuged at 8000g 4 °C for 10 minutes, take the supernatant into a new centrifuge tube and keep it on ice for detection.

2. For liquid samples
Detect directly.
V. ASSAY PROCEDURE

Add following reagents into the microplate:

<table>
<thead>
<tr>
<th>Reagent</th>
<th>Sample</th>
<th>Control</th>
<th>Standard</th>
<th>Blank</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sample</td>
<td>20 μl</td>
<td>20 μl</td>
<td>--</td>
<td>--</td>
</tr>
<tr>
<td>Standard Solution</td>
<td>--</td>
<td>--</td>
<td>20 μl</td>
<td>--</td>
</tr>
<tr>
<td>Distilled water</td>
<td>--</td>
<td>80 μl</td>
<td>--</td>
<td>20 μl</td>
</tr>
<tr>
<td>Substrate I</td>
<td>60 μl</td>
<td>--</td>
<td>60 μl</td>
<td>60 μl</td>
</tr>
<tr>
<td>Substrate II</td>
<td>20 μl</td>
<td>--</td>
<td>20 μl</td>
<td>20 μl</td>
</tr>
</tbody>
</table>

Mix, put it in the oven, 37 °C for 30 minutes.

| Dye Reagent I   | 50 μl  | 50 μl   | 50 μl    | 50 μl |
| Dye Reagent II  | 50 μl  | 50 μl   | 50 μl    | 50 μl |

Mix, wait for 20 minutes, record absorbance measured at 540 nm.
VI.  CALCULATION

**Unit Definition:** One unit of NR activity is defined as the enzyme generates 1 μmol of NO$_2^-$ per hour.

1. According to the protein concentration of sample

\[
NR \ (U/mg) = C_{\text{Standard}} \times \frac{(OD_{\text{Sample}} - OD_{\text{Control}})}{(OD_{\text{Standard}} - OD_{\text{Blank}})} \times \frac{V_{\text{Sample}}}{C_{\text{Protein}} \times V_{\text{Sample}}} \times \frac{V_{\text{Sample}}}{T} \\
= 20 \times \frac{(OD_{\text{Sample}} - OD_{\text{Control}})}{(OD_{\text{Standard}} - OD_{\text{Blank}})} \times \frac{V_{\text{Sample}}}{C_{\text{Protein}}} 
\]

2. According to the weight of sample

\[
NR \ (U/g) = C_{\text{Standard}} \times \frac{(OD_{\text{Sample}} - OD_{\text{Control}})}{(OD_{\text{Standard}} - OD_{\text{Blank}})} \times \frac{V_{\text{Sample}}}{W \times V_{\text{Assay}}} \times \frac{V_{\text{Sample}}}{V_{\text{Assay}}} \times \frac{1}{T} \\
= 20 \times \frac{(OD_{\text{Sample}} - OD_{\text{Control}})}{(OD_{\text{Standard}} - OD_{\text{Blank}})} \times \frac{1}{W} 
\]

*C$_{\text{Standard}}$: the protein concentration, 10 μmol/ml;
*C$_{\text{Protein}}$: the protein concentration, mg/ml;
*W*: the weight of sample, g;
*V$_{\text{Sample}}$: the volume of sample, 0.02 ml;
*V$_{\text{Assay}}$: the volume of Assay buffer, 1 ml;
*T*: the reaction time, 0.5 hour.
VII. TYPICAL DATA

The standard curve is for demonstration only. A standard curve must be run with each assay.

Detection Range: 0.1 μmol/ml - 10 μmol/ml

VIII. TECHNICAL SUPPORT

For troubleshooting, information or assistance, please go online to www.cohesionbio.com or contact us at techsupport@cohesionbio.com

IX. NOTES